

## EFFECT OF PEG STRESS ON GROWTH AND ASSOCIATED PHYSIOLOGICAL AND BIOCHEMICAL CHANGES IN SELECTED AND NON SELECTED UPLAND RICE

P. CHANDRASEKHARA REDDY\*, G.K. HALESH AND S.N. VAJRANABHAIAH

Department of Crop Physiology, UAS, GKVK, Bangalore - 560 065, Karnataka, India

Received on 7 May, 2003, Revised on 20 Dec., 2004

### SUMMARY

Calli of two rice genotypes, viz. Rasi and Valya were selected under PEG stress. Both varieties exhibited significant differences in growth, turgor potential, solute accumulation and lipid peroxidation parameters. There was better growth, increased better turgor potential values, greater accumulation of reducing sugars, amino acids, proline and superoxide dismutase (SOD) activity in the selected tissues of the varieties. However, between the two genotypes, tissues of Rasi exhibited higher values in terms of growth, water potential, solute accumulation and SOD than cv Valya which exhibited lowest values. In this study, it was concluded that the recurrent selection has improved the PEG tolerance in Rasi than Valya yet normal growth was not restored as that of control genotypes.

**Key words:** PEG stress, pressure potential, solute potential, superoxide dismutase, water potential.

### INTRODUCTION

Cellular mechanism of stress tolerance are important in developing drought tolerance among rainfed crops. Somaclonal selection technique is one of the recent methods applied to develop tolerant crops. (Warne and Hickok 1987). *In vitro* selection to develop stress tolerance rainfed rice is of utmost importance (Kavikishor and Reddy 1984, Chandrasekhara Reddy *et al.* 1994). Accumulation of different solutes in rice varied under water stress; composition of solute concentration in stressed cells accumulated more sucrose and reducing sugars and exhibited greater osmotic potential, than control (Chandrasekhara Reddy *et al.* 1994).

Growth of non adapted or adapted calli in stress media is shown to have different mechanisms of tolerance, developed during adaptation and selection. Generally, the

phenomenon of growth is accompanied by changing water relations and osmotic adjustments leading to build up of turgor pressure, reaction of more positive turgor to the continued extension of cell wall and growth (Hasegawa *et al.* 1984).

Drought/ stress affects physiological processes both at whole plant and cellular levels (Morgan 1984). Stress affects the membrane integrity (Dhindsa and Matowe 1981). The activity of some of the enzymes leads to a greater production of free radicals. These free radicals cause lipid peroxidation and membrane deterioration in plants (Chandrasekhara Reddy and Vajranabhaiah 1993). These lipid peroxidation products inhibit the protein synthesis by hydrolysis of m-RNA (Corcuera *et al.* 1989). The mechanism of prevention of lipid peroxidation consists of free radical scavenging enzymes, including superoxide dismutase and catalase which scavenge the free radicals,

\* Corresponding author

thus lead to drought tolerance (Spsychalla and Desborough 1990). Hence, in the present study, cell lines selected under PEG stress are compared with the non selected ones to understand the cellular mechanisms of drought endurance as a step towards development of improved rice genotypes.

## MATERIALS AND METHODS

Seed (embryo) calli from two rice genotypes, viz. Rasi and Valya initiated on Murashige and Skoog's (1962) modified medium supplemented with 2, 4-D (2 mg l<sup>-1</sup>) and kinetin (0.25 mg l<sup>-1</sup>) were subcultured and maintained on the same medium, where NAA (2.0 mg l<sup>-1</sup>) replaced 2, 4-D.

Polyethylene glycol (PEG 6000) was incorporated in the media to create -0.4, -0.6, -0.8 and -1.0 Mpa of stress. Callus initiated in these were cultured repeatedly in PEG free and PEG MS media with growth regulators (NAA 2.0 mg l<sup>-1</sup> and kinetin 0.25 mg l<sup>-1</sup>) for nine generations (R9), were used for this experiment. Calli

grown on PEG free and PEG medium from R10 generation, the fresh (FW) and dry (DW) weights and cell number per culture were determined on 30<sup>th</sup> day after inoculation (DAI). Water potential (-Ψ<sub>w</sub>) was determined as described earlier (Barr's 1968), solute potential (-Ψ<sub>s</sub>) was determined by using vapour pressure osmometer (Wescor, HP-115) and pressure potential (+Ψ<sub>p</sub>) was calculated as difference between -Ψ<sub>w</sub> and -Ψ<sub>s</sub> (+Ψ<sub>p</sub> = -Ψ<sub>w</sub> - (-Ψ<sub>s</sub>)). Solutes, reducing sugars (Nelson 1994), proline (Bates *et al.* 1973), amino acids (Spices 1957) and melon dialdehyde (MDA) (Chandrashekhara Reddy and Vajranabhaiah 1993), superoxide dismutase (SOD) (Beauchamp and Fridowich 1971) and soluble proteins (Lowry *et al.* 1951) were also determined.

## RESULTS AND DISCUSSION

Fresh (FW) and dry (DW) weights reduced by PEG stress more at higher levels of stress. This decline was common in both the genotypes. Selected tissues (ST) maintained higher growth, as indicated in FW and DW recorded on 30<sup>th</sup> day at all stress levels, than in

**Table 1.** Effect of PEG stress on selected and non selected rice calli

| Treatments<br>genotypes | FW (mg) |       | DW (mg) |      | Cell no (x 10 <sup>7</sup> ) |      |
|-------------------------|---------|-------|---------|------|------------------------------|------|
|                         | ST*     | NST** | ST      | NST  | ST                           | NST  |
| CONTROL                 |         |       |         |      |                              |      |
| Rasi                    | 647.0   |       | 75.8    |      | 2.30                         |      |
| Valya                   | 569.0   |       | 58.3    |      | 1.50                         |      |
| STRESS (-Mpa)           |         |       |         |      |                              |      |
| Rasi                    |         |       |         |      |                              |      |
| 0.4                     | 361     | 186   | 67.4    | 22.4 | 1.66                         | 1.16 |
| 0.6                     | 262     | 162   | 31.3    | 19.9 | 1.50                         | 1.05 |
| 0.8                     | 202     | 131   | 24.0    | 16.1 | 1.29                         | 0.83 |
| 1.0                     | 152     | 91    | 16.4    | 11.6 | 1.04                         | 0.58 |
| Valya                   |         |       |         |      |                              |      |
| 0.4                     | 193     | 158   | 20.9    | 16.0 | 1.10                         | 1.00 |
| 0.6                     | 149     | 135   | 16.4    | 14.1 | 0.90                         | 0.70 |
| 0.8                     | 125     | 81    | 14.0    | 8.7  | 0.50                         | 0.30 |
| 1                       | 118     | Dead  | 13.0    | Dead | 0.50                         | Dead |
| CD 0.01 Var             | 36      | 30    | 3.00    | 2.20 | 0.10                         | 0.10 |

\*Selected tissue - tissues continuously grown under PEG stress media for 10 generations

\*\* Non-selected tissues - tissues continuously grown in control medium and transferred to stress medium on 10<sup>th</sup> generation.

## EFFECT OF PEG STRESS ON RICE CALLI

corresponding treatments given to non-selected (NST) calli of same age group. The same observations were made for cell numbers also, where decrease in cell number due to increase in PEG stress led to more reduction in NST than ST (Table 1). Water ( $-\Psi_w$ ), solute ( $-\Psi_s$ ) and pressure ( $+\Psi_p$ ) potentials (Table 2) examined during the

course of 30 DAI in culture revealed that the  $-\Psi_w$  and  $-\Psi_s$  were more negative in ST in general at all stress level and ST of rice genotypes exhibited greater reduction in  $-\Psi_w$  and  $-\Psi_s$  than NST at all stress levels (Table 2). Pressure potential values were also generally more positive in ST of all the genotypes than NST. A decline in  $+\Psi_p$  with

**Table 2.** Effect of PEG stress on water relation (-Mpa) in selected and non selected rice calli

| Treatments<br>genotypes | $-\Psi_w$ |      | $-\Psi_s$ |      | $+\Psi_p$ |      |
|-------------------------|-----------|------|-----------|------|-----------|------|
|                         | ST        | NST  | ST        | NST  | ST        | NST  |
| CONTROL                 |           |      |           |      |           |      |
| Rasi                    | 0.80      |      | 1.79      |      | 0.99      |      |
| Valya                   | 0.80      |      | 1.29      |      | 0.49      |      |
| STRESS (-Mpa)           |           |      |           |      |           |      |
| Rasi                    |           |      |           |      |           |      |
| 0.4                     | 2.07      | 1.47 | 2.96      | 2.23 | 0.89      | 0.76 |
| 0.6                     | 2.67      | 1.90 | 3.40      | 2.51 | 0.73      | 0.61 |
| 0.8                     | 3.07      | 2.30 | 3.90      | 2.67 | 0.83      | 0.37 |
| 1.0                     | 3.37      | 2.73 | 4.14      | 2.97 | 0.77      | 0.23 |
| Valya                   |           |      |           |      |           |      |
| 0.4                     | 1.83      | 1.57 | 2.27      | 1.60 | 0.44      | 0.03 |
| 0.6                     | 2.50      | 1.83 | 2.55      | 1.88 | 0.05      | 0.05 |
| 0.8                     | 2.50      | 1.40 | 2.59      | 1.33 | 0.09      | 0.07 |
| 1.0                     | 2.90      | Dead | 2.90      | Dead | 0.0       | Dead |
| CD 0.01 Var             | 0.15      | 0.11 | 0.16      | 0.13 | 0.09      | 0.08 |

**Table 3.** Effect of PEG stress on solute accumulation in selected and non selected rice calli

| Treatments<br>genotypes | Reducing sugars (mM) |      | Amino Acid (mM) |      | Proline (mM) |      |
|-------------------------|----------------------|------|-----------------|------|--------------|------|
|                         | ST                   | NST  | ST              | NST  | ST           | NST  |
| CONTROL                 |                      |      |                 |      |              |      |
| Rasi                    | 177                  |      | 170             |      | 54           |      |
| Valya                   | 116                  |      | 128             |      | 42           |      |
| STRESS (-Mpa)           |                      |      |                 |      |              |      |
| Rasi                    |                      |      |                 |      |              |      |
| 0.4                     | 204                  | 110  | 185             | 179  | 91           | 66   |
| 0.6                     | 245                  | 189  | 196             | 183  | 115          | 78   |
| 0.8                     | 276                  | 200  | 202             | 184  | 166          | 86   |
| 1.0                     | 250                  | 210  | 213             | 180  | 187          | 92   |
| Valya                   |                      |      |                 |      |              |      |
| 0.4                     | 119                  | 129  | 130             | 116  | 69           | 47   |
| 0.6                     | 130                  | 115  | 141             | 100  | 98           | 35   |
| 0.8                     | 121                  | 77   | 131             | 26   | 97           | 41   |
| 1.0                     | 107                  | Dead | 118             | Dead | 94           | Dead |
| CD 0.01                 | 17                   | 16   | 8.0             | 6.0  | 5.0          | 4.0  |

increase in PEG concentration was noticed among the NST calli (Table 2). Solutes, reducing sugar (RS), amino acid (AA) and proline accumulation in general are greater in the ST than in NST at all the stress levels (Table 3).

Two genotypes, Rasi and Valya, differed significantly in their growth, solute accumulation and also water relations. In general Rasi performed far better in terms of its growth, solute accumulation and osmotic adjustment than Valya (Table 1, 2 and 3) in all the stress levels.

Selected (S-S and S-C) tissues which maintained higher growth contained lower MDA levels under stress (Table 4). There was a general increase in MDA contents of both ST (S-S) and NST (C-S) tissues of all the rice varieties when subjected to stress. But this enhancement was relatively low in ST (S-S). SOD activity in the control tissues grown in normal medium was low. The activity of SOD increased, when the control tissues were subjected to stress. ST in PEG and PEG free media exhibited higher values of SOD activity. Highest activity was found in ST grown in stress medium. ST in stress medium maintained relatively higher protein than NST subjected to stress. Protein contents further increased in ST (S-S) when transferred to normal medium (S-C).

Rasi and Valya were on two extremes in their accumulation of MDA, SOD and soluble proteins. Rasi exhibited lower MDA and higher SOD activity and protein content as compared to Valya in all situations (Table 4).

Continued decline in FW and DW with increasing stress is common to both ST and NST tissues. Better growth, in terms of FW and DW of the ST than NST at 10<sup>th</sup> generation (R10) shows that capacity to tolerate stress is more in ST rather than NST. NST of Valya became senescent at 1.0 Mpa of stress because of highly susceptible nature of stress. Lower percentage reduction in growth among the ST calli at different stress levels over the NST, clearly suggests that selection enhanced the tolerance and growth. Similar observations were reported earlier (Bressan *et al.* 1981). However in ST the growth is not fully recovered when they were transferred to control medium. This pattern of response is in conformity with that of tomato (Handa *et al.* 1983). Effect of stress has reduced proportionately the cell number per culture, suggesting a retardation effect of stress on cell division too. Similar observation made by Chandrasekhara Reddy and Vajranabhaiah (1996) in rice and this cell division is less affected in ST than NST at all stress level. This suggests that the selected tissues are able to survive and grow better under stress. Cell number was greatly affected in Valya as compared to Rasi at all stress levels this

**Table 4.** Effect of PEG stress on MDA content, SOD activity and soluble proteins in ST and NST of rice calli

| Treatments   | Fw (mg) | Dw (mg) | MDA (OD/g dw) | SOD (Units/mg dw) | Sol-proteins (mg/g dw) |
|--------------|---------|---------|---------------|-------------------|------------------------|
| <b>Rasi</b>  |         |         |               |                   |                        |
| C            | 657     | 77      | 1.52          | 145               | 50                     |
| C-S          | 161     | 22      | 2.61          | 278               | 43                     |
| S-C          | 459     | 68      | 1.66          | 261               | 48                     |
| S-S          | 201     | 28      | 2.04          | 373               | 47                     |
| <b>Valya</b> |         |         |               |                   |                        |
| C            | 569     | 58      | 3.00          | 266               | 60                     |
| C-S          | 118     | 13      | 5.14          | 250               | 37                     |
| S-C          | 301     | 31      | 4.00          | 261               | 50                     |
| S-S          | 128     | 15      | 5.00          | 252.0             | 43                     |
| CD 0.01      | 10      | 2       | 0.1           | 10.0              | 2.0                    |

C-Control

C-S - Control to stress medium (non-selected)

S-C - Stress medium to control medium (adapted cells in control medium)

S-S - Stress to stress medium (selected)

indicated that Rasi had more osmotic adjustment than Valya which led to better growth under severe stress.

Continued selection under stress from  $R_1$  through  $R_{10}$  has led to the amplification of responses by way of greater reduction in  $-\Psi_w$  and  $-\Psi_s$  leading to more positive turgor at  $R_{10}$  in tissues of all genotypes and at different stress levels. This osmotic adjustment leading to greater growth and turgor in ST than NST grown at different concentration of PEG are also reported earlier (Handa et al. 1986). There are identical reports on growth and osmotic adjustments in ST and NST of brassica (Chandler and Thorpe 1987, Paek et al. 1988). Selected tissues showed the degree of inhibition of growth to be lesser than NST under decreasing  $-\Psi_w$ . Similar observation made earlier (Fallon and Philips 1989). Where the decline in FW and DW continued in un-adapted cells with decrease in  $-\Psi_w$ . This was greater than adapted cells.

A comparison of the ST and NST revealed that the degree of response is greater in ST calli which accumulated more of all the three solutes. Similar response was reported earlier (Fallon and Philips 1989), where, greater accumulation of the solute was of higher order than present observation in ST. Accumulation of these solutes (RS, AA and proline) in the ST and NST cell lines under PEG stress indicates that this may be the part of osmotic adjustment mechanism operating to maintain the turgor.

Since, stress is also known to cause membrane damage and leakage of solutes, attempts were also made to study the extent of lipid peroxidation and defense mechanisms as an indication of drought tolerance. The MDA content may indicate extent of resistance to membrane lipid peroxidation, which is also important in stress tolerance. Desiccation tolerance in the ST have more active free radical scavenging enzyme systems which retard the peroxidation of membrane lipids and accumulation of MDA. The free radicals quenching system is represented by SOD enzyme, enhanced activity of this enzyme in all the ST suggests prevention of damage caused to cell membrane by scavenging super oxide radicals. The increase in specific activity of this enzyme in the light of the reduced soluble protein contents in the ST and NST subjected to stress and ST in normal medium suggests more of biosynthesis of this class of proteins. This means the switching on and off of defensive mechanism against enhanced lipid peroxidation

caused by stress. Such similar defensive mechanisms have been reported earlier (Spsychalla and Desborough 1990).

## REFERENCES

- Barrs, H.D. (1968) Determination of water deficits in plant tissues. In: T.E. Kozlowski (ed.), *Water Deficits and Plant Growth* vol I: Acad press, New York.
- Bates, L.S., Waldren, R.P. and Teare, I.D. (1973), Rapid determination of free proline in water stress studies. *Plant & Soil*. **39**: 205-208.
- Beauhamp, D.C. and Fridovich, I. (1971). Superoxide dismutase; improved assays and an assay applicable to acrylamide gels. *Anal. Biochem*. **44**: 276-287.
- Bressan, R.A., Hasegawa, P.M. and Handa, A.K., (1981). Resistance of cultured higher plant cells to polyethylene glycol-induced water stress. *Plant Sci*. **21**: 23-30.
- Chandler, S.F. and Thorpe, T.A., (1987). Characterization of growth, water relations and proline accumulation in sodium sulphate tolerant callus of *Brassica napus* L. cv Weston (Canola). *Plant Physiol*. **84**: 106-111.
- Chandrasekhara Reddy, P., Vajranabhaiah, S.N. and Prakash, A.H. (1994). Varietal responses of upland rice calli to polyethylene glycol (PEG 6000) stress. *Adv Plant Sci*. **7**: 12-17.
- Chandrasekhara Reddy, P. and Vajranabhaiah, S.N. (1996).  $^{14}C$  sucrose uptake and incorporation in stress tolerant calli of upland rice (*Oryza sativa* L.). *Adv Pl. Sci*. **4**: 61-65.
- Chandrashekhara Reddy, P. and Vajranabhaiah, S.N. (1993). Drought induced lipid peroxidation : Defensive mechanisms in upland rice (*Oryza sativa* L.) seeds during germination. *Adv. Plant Sci*. **6**: 229-236.
- Corcuera, L.J., Hintz, M. and Pahlich, E. (1989). Effect of polyethylene glycol on protein extraction and enzyme activities in potato cell cultures. *Phytochem*. **28**: 1569-1591.
- Dhindsa, R.S. and Matowe, W. (1981). Drought tolerance in two mosses correlated with enzymatic defense against lipid peroxidation. *J. Exp. Bot*. **32**: 79-91.
- Fallon, K.M. and Philips, R. (1989). Response of water stress in adapted and non adapted carrot cell suspension cultures. *J. Exp. Bot*. **40**: 681-687.

- Handa, S., Bressan, R.A., Handa, A.K., Carpita, N.C. and Hasegawa, P.M. (1983). Soluble contribution to osmotic adjustments in cultured plant cells adapted to water stress. *Plant Physiol.* **73**: 834-843.
- Handa, S., Handa, A.K., Hasegawa, P.M. and Bressan, R.A. (1986). Proline accumulation and the adaptation of cultured plants cells to water stress. *Plant Physiol.* **80**: 938-945.
- Hasegawa, P.M., Bressan, R.A., Handa, A.K. and Handa, S. (1984). Cellular mechanisms of tolerance to water stress, *HortSci.* **19**: 7-13.
- Kavikishor, P.B. and Reddy, G.M. (1984). *In vitro* selection of PEG and NaCl resistance in rice. Mutation breeding news letter, no. 24, 6 (edn). Dept Genetic. Osmania University, Hyderabad, Andhra Pradesh, India.
- Lowry, D.H. Roseborough, N.T., Farr, A.L. and Randall, R.J. (1951). Protein measurement with folin phenol reagents. *J. Biol. Chem.* **193**: 265-275.
- Morgan, J.M. (1984). Osmoregulation and water stress in higher plants. *Ann Rev. Plant Physiol.* **35**: 299-319.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bio-assays with tobacco tissue cultures. *Physiol. Plant.* **15**: 473-477.
- Nelson, N. (1944). A photometric adaptation of the somogyi method for determination of glucose. *J. Biol. Chem.* **153**: 375-380.
- Paek, K. Y., Chadler, S.F. and Thorpe, T.A. (1988). Physiological effects of Na<sub>2</sub>SO<sub>4</sub> and NaCl on callus cultures of *Brassica campestris* (Chinese cabbage). *Physiol. Plant.* **72**: 160-166.
- Spices, J.R. (1957) Ninhydrin method colorimetric procedure for amino acids. *Methods in Enzymology.* **3**: 468-471.
- Spychalla, J.P. and Desborough, S.L. (1990) Superoxide dismutase, catalase and tocopherol contents of stored potato tubers. *Plant Physiol.* **44**: 1214-1218.
- Warne, T.R. and Hickok, L.G. (1987) Single gene mutants tolerance to NaCl in the fern *Ceratopteris*: characterization, genetic analysis, *Plant Sci.* **52**: 49-55.